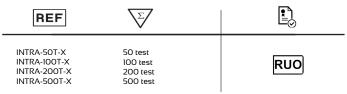
## INTRACELL

### Fixation and permeabilization kit for Flow Cytometry





### 1. INTRODUCTION

Staining cells for internal antigens techniques have been devised for permeabilizing cells so that specific antibody can diffuse in and out and access internal antigens. Briefly, cells are fixed with formaldehyde to preserve their morphology, and saponin, a plant glycoside and mild nonionic detergent, is used to generate pores of approximately 8nm in the membrane which allow molecules of up to 200 RDa to pass through. Because of slow diffusion in and out of the cell, longer staining and washing times are required, cell and antibody preparations must be of high quality, and antibody

must be at optimal titre to reduce nonspecific background.

Other treatments can be used. For staining nucleic acid, fixation and permeabilization with alcohol and acetic acid reduces degradation of RNA and DNA. Cells may also be stained with antibody to a cell surface antigen for multicolour analysis.

IntraCell is intended for fixation and permeabilization of cells in suspension for flow cytometry analysis.Immunological detection studies of intracellular antigens on structures such as cytoplasmic and/or nuclear enzymes, requires the permeabilization of the cell membrane in order to allow interaction of the antibody with its intended target. In order to allow recognizable cellular integrity after permeabilization a fixation step involving cross-linking or denaturation is required. This is usually accomplished by aldehyde or alcohol fixation followed by detergent permeabilization of the cell membrane. Labeling of cell surface antigens with antibodies prior to the fixation step is possible thus allowing simultaneous surface antigens phenotyping with internal antigen expression in multiparameter fluorescent cytometric analysis.

IntraCell is developed for use with all flow cytometers and reagents remains intact the cell surface marker expression and the cell properties of FSC and SSC.

### 2. MATERIALS PROVIDED

Reagent A: Fixative solution

Reagent B: Permeabilization solution

#### 3. MATERIALS DON'T PROVIDED

PBS Working Solution. IX Phosphate- buffered saline solution (PBS) + 0,1 NaN3 + 4% BSA.

### 4. APPROPRIATE STORAGE AND HANDLING CONDITIONS

Store in the dark, refrigerated between 2 °C and 8 °C. DO NOT FREEZE. The antibody is stable until the expiry date stated on the vial label if kept at 2°C-8°C. Do not use after the dateindicated.

Once the vial is open, the product is stable for 90 days.

### 5. EVIDENCE OF DETERIORATION

Reagents should not be used if any evidence of deterioration is observed. For more information, please contact our technical service: tech@immunostep.com

The product's normal appearance is a semi-transparent, colourless liquid. It should not be used if liquid medium is cloudy or contains precipitate. It should be odourless.

#### 6. WARRANTY

Warranted only to conform to the quantity and contents stated on the label or in the product labelling at the time of delivery to the customer. Immunostep disclaims hereby other warranties. Immunostep's sole liability is limited to either the replacement of the products or refund of the purchase price

### 7. RECOMMENDATIONS AND WARNINGS

- a) The reagents contain sodium azide. In acid conditions, it is transformed into hydrazoic acid, a highly toxic compound. Azide compounds must be diluted in running water before being discarded. These conditions are recommended so as to avoid deposits in plumbing, where explosive conditions could develop. The safety data sheet (SDS) is available online at www.immunostep.com
- b) Avoid microbial contamination of the reagent.
- c) Protect from light. Use dim light during handling, incubation with cells and prior to analysis.
- d) Never mouth pipette.
- e) In the case of contact with skin, wash in plenty of water.
- The samples should be handled in the same way as those capable of transmitting infection. Appropriate handling procedures should be guaranteed.
- g) Do not use after the expiry date indicated on the vial.
- h) Deviations from the recommended procedure could invalidate the analysis results.
- i) FOR IN VITRO DIAGNOSTIC USE.
- j) For professional use only.
- ${\bf k})$  Before acquiring the samples, it is necessary to make sure that the flow cytometer is calibrated and compensated.

# 8. PROTOCOL FOR SAMPLE PREPARATION AND INTRACELLULAR STAINING

- 1. Pipette 50  $\mu l$  of cell suspension to be analysed (up to 106 cells) into each tube.
- For each sample, add an appropriate volume of conjugated antibody directed to the cell surface antigen of interest and the appropriate isotype control. Incubate for I5 minutes in the dark at room temperature. (This step is only necessary is you want to perform a direct immunofluorescence staining for a cell surface antigen)
- 3. Add 100 µl of IntraCell Reagent A, (Fixative), to each tube. Mix gently.
- 4. Incubate for 15 minutes at room temperature.
- 5. Wash once in 2 ml PBS Working Solution 1X.
- 6. Centrifuge for 5 minutes at 300 xg, then aspirate supernatant, leaving approximately 50  $\,\mu$ l of fluid and vortex to ensure that the cell pellet are in suspension.
- Add 100 µl of IntraCell Reagent B (Permeabilization), to each tube. Add the appropriate volume of conjugated intracellular antibody specific for the intracellular antigen and the appropriate isotype control.
- 8. Incubate for 15 minutes in the dark at room temperature.
- 9. Wash once in 2 ml PBS Working Solution IX.
- 10. Centrifuge for 5 minutes at 300 xg, then aspirate supernatant, leaving approximately 50 $\mu$ l of fluid  $\,$  and vortex to ensure that the cell pellet are in suspension.
- Resuspend the cell pellet in 0,5 ml of 1% paraformaldehyde solution or an appropriate fluid for flow cytometric use, and store in the dark at 2-8 °C. Fixed cells should be analyzed within 24hours.

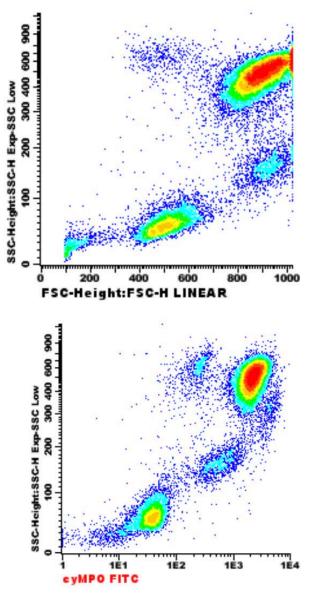


Figure 1. Intracellular FITC mouse anti-human myeloperoxidase (MPO) vs side (SSC) scatter of a normal blood sample.

#### 9 **INTRACELLULAR PROTEINS EVALUATED**

Enzymes	Myeloperoxidase, Carboxipeptidase, Granzyme B <sup>3</sup> , Perforine <sup>3</sup>
Cytoplasmatic CD molecules	CD3, CD13, CD22, CD62P, CD63, CD68, CD79a
Nuclear proliferation markers	BrdU, Ki-67 <sup>7</sup> , PCNA, TdT
Oncoproteins	Bcl-2, c-Myc, p53 <sup>9</sup>
Cytokeratines	CK19 <sup>6</sup>
Caspases	CASP3 <sup>8</sup>
Cytokines and cheokines	IFN-γ, TNF-α, IL1-β, IL-2, IL-4, IL- 10, IL-12, IL-13, IL-16, RANTES
Immunoglobulins	lgA, lgG, lgD, lgM, kappa, lambda
Other molecules	ZAP-70, cyclins, transfected cells, intracellular parasites <sup>4,5.7</sup> , Iamin A / C7, MDR (multidrug resistance) <sup>1</sup>

#### 10. SENSIBILITY

The quality of each batch of intracell is determined by fixation and permeabilization of several samples of peripheral blood from healthy donors with some of the markers mentioned above and the corresponding comparison of the characteristics of size and complexity of the preparation of leukocyte processed.

#### TROUBLESHOOTING 11.

- a) Most conjugated antibodies are suitable for use together with IntraCell. However, there are some antigen determinants that are sensitive to the fixation with formaldehyde. And optimal fixation time or formaldehyde concentration, should be determinate for each antibody conjugated.
- b) The fixation/permeabilization reagent (INTRACELL) contained paraformaldehide and their cross-linking propriety didn't allow RNA extraction.
- c) In the staining procedure, particular attention should be paid to the incubation times.

#### REFERENCES 12.

1. Castalta-Lopes J. et al. Efflux Pumps Modulation in Colorectal Adenocarcinoma Cell Lines: The Role of Nuclear Medicine. Journal of Cancer Therapy. Vol.2 No.3(2011), Article ID:6693. 2. Brito AF. Et al. Hepatocellular Carcinoma and Chemotherapy: The Role of p53. Chemotherapy 2012:58:381-386.

3. Alba Fernández-Sánchez, et al. DNA demethylation and histone H3K9 acetylation determine the active transcription of the NKG2D gene in human CD8+ T and NK cells. Epigenetics. 2013 Jan 1; 8(1): 66-78.

4. Nieves Ayllón, et al. Anaplasma phagocytophilum Inhibits Apoptosis and Promotes Cytoskeleton Rearrangement for Infection of Tick Cells. Infect Immun. 2013 Jul; 81(7): 2415-2425.

5. Victoria Naranjo, et al. Reciprocal Regulation of NF-kB (Relish) and Subolesin in the Tick Vector, Ixodes scapularis. PLoS One. 2013; 8(6): e65915.

6. Ángela Santoro, et al. Relationship between CKI9 expression, deregulation of normal keratinocyte differentiation pattern and high risk-human papilloma virus infection in oral and oropharyngeal squamous cell carcinoma. Infect Agent Cancer. 2015; 10: 46.

7. Margarita Villar, et al. Identification and Characterization of Anaplasma phagocytophilum Proteins Involved in Infection of the Tick Vector, Ixodes scapularis. PLoS One. 2015; 10(9): e0137237.

8. José Mendes, et al. L744,832 and Everolimus Induce Cytotoxic and Cytostatic Effects in Non-Hodgkin Lymphoma Cells. Pathology & Oncology Research, April 2016, Volume 22, Issue 2, pp 301-309.

9. Rocío Navarro, et al. Role of nucleotide bindingoligomerization domain 1 (NODI) in pericyte mediated vascular inflammation. J Cell Mol Med. 2016 May; 20(5): 980-986.

#### 13. SYMBOLS



14. MANUFACTURED BY



Immunostep

#### IMMUNOSTEP S.L. Address: Avda. Universidad de Coimbra, s/n

Cancer Research Center (C.LC) Campus de Unamuno 3700 7 Salamanca (Spain) Telf/fax: (+34) 923 294 827 E-mail: info@immunostep.com www.immunostep.com